Critical assessment of assembly strategies for non-model species mRNA-Seq data and application of next-generation sequencing to the comparison of C₃ and C₄ species

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Abstract

Next-generation sequencing enables the study of species without a sequenced genome at the ‘omics’ level. Custom transcriptome databases are generated and global expression profiles can be compared. However, the assembly of transcriptome sequence reads into contigs remains a daunting task. In this study, five different assembly programs, both traditional overlap-based, ‘read-centric’ assemblers and de Bruijn graph data structure-based assemblers, were compared. To this end, artificial read libraries with and without simulated sequencing errors were constructed from Arabidopsis thaliana, based on quantitative profiles of mature leaf tissue. The open source TGICL pipeline and the commercial CLC bio genomics workbench produced the best assemblies in terms of contig length, hybrid assemblies, redundancy reduction, and error tolerance. The mature leaf transcriptomes of the C₃ species Cleome spinosa and the C₄ species Cleome gynandra were assembled and analysed. The pathways and cellular processes tagged in the transcriptome assemblies reflect processes of a mature leaf. The databases are useful for extracting transcripts related to C₄ processes as full-length or nearly full-length sequences.

Key words: Assembly, C₄, next-generation sequencing, transcriptome.

Introduction

Sequence information, both qualitative (the sequence itself) and quantitative (how much each transcript is expressed), is important for the analysis of any trait at the molecular level. Next-generation sequencing (NGS) technologies have recently become widely available, and the creation of custom transcriptomes a possibility (Weber et al., 2007; Novaes et al., 2008; Alagna et al., 2009; Barakat et al., 2009; Dassanayake et al., 2009; Wang et al., 2009; Kumar and Blaxter, 2010). Using NGS to study the transcriptome of a species without a sequenced genome, such as a C₄ species other than Zea mays or Sorghum bicolor, simultaneously produces a transcriptome database for the tissue sampled as well as an expression profile of the tissue (Bräutigam et al., 2011). It is even possible to compare the expression profiles of two different species, for example a C₃ and a C₄ species, with one another (Bräutigam and Gowik, 2010; Bräutigam et al., 2011).

Different NGS technologies are currently available commercially (Metzker, 2010). Common to all available NGS technologies is that they produce much more sequence information compared with traditional Sanger sequencing at a much lower cost. However, there is no free lunch (yet), and with current technologies the payment is in short reads, from 36 bases (with Illumina technology; longer reads of up to 100 bases are possible at increased cost), over 75 bases (with SOLiD technology), and to ~450 bases (with Roche/454 technology). Roche/454 technology will produce fewer reads per run than Illumina and SOLiD, though. Although the sequence reads themselves are longer, the total sequence output is only about one-tenth of Illumina’s output and ~1/100th of SOLiD’s output. The possible applications of the long read and short read technologies in the context of plant research and their advantages and disadvantages have been reviewed in detail (Bräutigam and Gowik, 2010).
Briefly, in species without a sequenced genome, longer reads will facilitate both the assembly of a transcriptome database and the reliable quantification of expression. Therefore, the only technology which gives us reads >200 bases is currently used: Roche/454.

If transcriptome sequence information is generated for a species without a sequenced genome, two analyses are possible: the quantification of expression by aligning (also referred to as mapping) the reads to a related reference genome as explored by Bräutigam et al. (2011) and the assembly of the transcriptome to provide qualitative sequence information. A read mapping is prone to errors if the reads do not exactly match the reference (Palmieri and Schlotterer, 2009), a caveat certainly true for mapping only to a related reference genome and not the genome itself. Longer reads ensure a more accurate mapping (Palmieri and Schlotterer, 2009). BLAT has been shown to map reads reliably when the method was applied to compare the expression profiles of a C3 and a C4 species (Schlotterer, 2009), a caveat certainly true for mapping only to a related reference genome and not the genome itself. Longer reads ensure a more accurate mapping (Palmieri and Schlotterer, 2009). BLAT has been shown to map reads reliably when the method was applied to compare the expression profiles of a C3 and a C4 species (Schlotterer, 2009). BLAT has been shown to map reads reliably when the method was applied to compare the expression profiles of a C3 and a C4 species (Schlotterer, 2009). BLAT has been shown to map reads reliably when the method was applied to compare the expression profiles of a C3 and a C4 species (Schlotterer, 2009).

In the quantitative comparison of the Cleome species C. gynandra and C. spinosa, the steady-state transcript levels of genes associated with C4 photosynthesis were increased between 20- and 250-fold in the C4 species, with the exception of malate dehydrogenase. This global approach also identified candidate genes for C4-related processes, such as intra- and intercellular metabolic transport, as well as candidates for regulators, which maintain the C4 state in mature leaves. Moreover, genes for protein biosynthesis, such as genes encoding ribosomal proteins, were more frequently down-regulated in the C4 species, as were many of the genes encoding Calvin–Benson cycle and photorespiratory enzymes, indicating that down-regulation of ribosomal proteins in the cytosol and chloroplast may be contributing to nitrogen efficiency in some C4 species.

However, the assembly of NGS reads remains a challenge. This is especially true for highly dynamic transcriptome read libraries. In principle, two different types of assemblers are available: a ‘read-centric’ overlap-based assembler, which has been used for assembling Sanger sequences, and an assembler specifically developed for handling the large amounts of reads provided by NGS, which is based on de Bruijn graph data structures (Flick and Birney, 2009). However, the new type of assemblers have been developed for assembling genomic rather than transcriptomic sequence libraries (Flick and Birney, 2009). While transcriptome libraries have dynamic ranges of 5-6 orders of magnitude between the highest abundant and the lowest abundant transcripts and their reads (Bräutigam et al., 2011), genomic libraries ideally have no dynamic range. Traditional assemblers include CAP3 (Huang and Madan, 1999), TGICL, which is a pipeline of a megablast-like tool connected to the CAP3s clustering algorithm (Pertea et al., 2003), and MIRA (Chevreux et al., 2004). New assemblers of the de Bruijn graph type are, for example, SOAPdenovo (http://soap.genomics.org.cn/soapdenovo.html) and Velvet (Zerbino and Birney, 2008). Commercial programs such as the CLC bio genomics workbench do not fully disclose the type of assembler integrated into the program. In plants, several attempts to reconstruct a plant transcriptome from NGS reads have been published with different assembly programs without any tests of whether one assembler outperformed any other (Alagna et al., 2009; Barakat et al., 2009; Wang et al., 2009). Such a critical assessment of assembler performance and suitability is conducted in this study.

The C4 syndrome, a complex trait evolved to concentrate carbon in the vicinity of Rubisco, has originated in >45 plant lineages in a striking example of convergent evolution (Sage, 2004). It serves to minimize the oxygenation reaction of Rubisco while maximizing CO2 fixation. C4 plants thus are either able to accumulate biomass much faster than plants without this carbon concentration mechanism (e.g. Z. mays or S. bicolor) or able to live in adverse conditions that minimize CO2 availability to and fixation by Rubisco, such as water limitation, heat, or poor soil conditions (Sage, 2004).

C4 plants have a biochemical CO2 pump: CO2 is fixed by phosphoenolpyruvate carboxylase (PEPC), an enzyme insensitive to O2 but with a higher affinity for HCO3-, in the mesophyll cells, whilst RuBisCO is localized to the bundle sheath cells. Several different enzymes and transport proteins transfer the CO2 as an acid with four carbon atoms, decarboxylate it to release the CO2 in the vicinity of RuBisCO, return a C4 acid to the site of PEPC, and regenerate the CO2 acceptor (Hatch, 1987). While the fixation of CO2 using the acceptor phosphoenolpyruvate (PEP) is always accomplished by the same enzyme, the C4 transfer acids can be malate and/or aspartate and the C3 transfer acid can be pyruvate, alanine, or PEP. Three different decarboxylation enzymes liberate the CO2, NAD-dependent malic enzyme (NAD-ME), NADP-dependent malic enzyme (NADP-ME), and PEP carboxykinase (PEPCK) (Hatch, 1987). The spatial separation of initial and final carbon fixation may be concomitant with the spatial separation of other anabolic pathways such as nitrogen and sulphur assimilation (Majeran et al., 2005) as well as limited oxygen production at the site of RuBisCO and increased ATP production (Meierhoff and Westhoff, 1993). In addition to this biochemical CO2 pump, several adaptations on the cellular and tissue levels are necessary (Hatch, 1987). The majority of C4 species, in terms of both species number and contribution to global biomass production, spatially separate RubisCO and PEPC in two different cell types called the mesophyll and the bundle sheath. For efficient C4 photosynthesis, CO2 released in the vicinity of RubisCO must not leak out of cells. A barrier is established either by a cell wall reinforced with lignin and/or suberin (Evert et al., 1977) or by positioning the RubisCO-containing chloroplasts ‘in the way’ of loss by diffusion (Muhaidat et al., 2007). While diffusion of CO2 must be prevented, diffusion of the transfer acids must not only be allowed, but must also be very effective to accommodate the immense metabolite flux through the C4 cycle, which operates at or above the speed of carbon fixation (Laisk and Edwards, 2000; Weber and von Caemmerer, 2010). Although all enzymes required for the C4 cycle are characterized at least
at the biochemical level and many also at the molecular level (Hatch, 1987), the majority of molecular changes underlying the tissue and cellular adaptations, the intra-
(Bräutigam et al., 2008; Majeran et al., 2008) and intercellular transport processes, such as plasmodesmatal regulation (Bothia, 1992; Sowinski et al., 2008), and most of the regulatory changes are unknown.

Forty-five origins of C₄ photosynthesis (Sage, 2004) provide at least 45 possible contrasting pairs of C₃ and C₄ plants to study. This study focuses on the C₄ plant C. gynandra (spider wisp, also known as ‘African cabbage’) and a C₃ relative, C. spinosa (spider plant). Cleome gynandra is currently known as the most closely related C₄ plant to Arabidopsis thaliana (thale cress) (Brown et al., 2005). It is a leafy annual plant from the African continent and forms part of the diet in African countries (van Rensburg et al., 2004). Within the genus Cleomeae, there are other species that show characteristics of C₄ plants, such as carbon isotope discrimination in, for example, C. angustifolia and C. oxalidae (Marshall et al., 2007). Since C. gynandra is easy to cultivate (it is considered an invasive weed in the USA; http://plants.usda.gov/java/profile?symbol=CLGY) and since it is a food plant for which seeds can be obtained in quantity from retailers, it is an attractive choice as an experimental organism. Transformation has recently been achieved (Newell et al., 2010). Cleome gynandra is an old world plant and probably a basal branch within the Cleomeae, although these basal branches are not well supported in phylogenetic trees (Inda et al., 2008).

A genome duplication event in the lineage of C. gynandra, which has 16 or 17 chromosomes, has been speculated about based solely on chromosome number (Inda et al., 2008). The choice for the C₃ plant in the comparison pair is not obvious. The basal branch of C. gynandra does not contain a C₃ relative and, based on the evolutionary trees available (most recent in Inda et al., 2008), the C₃ relatives are equidistant. The spider flower C. spinosa was chosen for its ease of cultivation and availability. Cleome spinosa is an ornamental plant which originated from South America. It is a member of the new world Cleomeae with a chromosome number of x=8 or 9 (Inda et al., 2008). Both plants can be cultivated alongside each other in a glass house or growth chamber. Under identical conditions in well-watered, rich soil, the C₃ plant will outgrow the C₄ plant. Extensive sequence information was not previously available for both species (Bräutigam et al., 2011).

To provide not only the quantitative information (Bräutigam et al., 2011) but also the best possible qualitative sequence information for the C₄ model C. gynandra, several different assemblers were tested in this study. To identify the most suitable assembler, a gold standard is needed against which the performance of the assemblers can be benchmarked. A simulated and therefore artificial read library, which represents the dynamics of a mature leaf transcriptome, was modelled based on the known A. thaliana genome. Assemblies were compared given that the ultimate best possible outcome, the real transcriptome, is known in this case. After the best assembler was determined, the sequence reads from both Cleome species were assembled, single nucleotide polymorphisms (SNPs) were annotated, and the transcript representation was analysed. The results of this study together with the recently published results of the transcriptome quantification (Bräutigam et al., 2011) enable the identification and study of transcripts involved in maintaining C₄ tissue and cell architecture, and regulating and executing C₄ photosynthesis in mature leaves.

**Materials and methods**

**Sequence read generation**

The sequence reads from C. gynandra and C. spinosa were generated as described in Bräutigam et al. (2011). They are available at NCBI’s short read archive under the accession numbers SRS002473 and SRS002474.

**Generation of the simulated read database**

Testing an assembly from sequencing reads that were generated from a species without a sequenced genome is problematic since the solution to the assembly, the correct transcriptome, is unknown. On the other hand, producing sequencing reads from a species with a sequenced genome for the express purpose of testing assembly programs is prohibitively expensive. To overcome these limitations, the quantitative information generated in Bräutigam et al. (2011) was used. If a Perl script draws the number of reads determined in Bräutigam et al. (2011) from each transcript and randomly distributes the reads along the length of the transcript, a simulated read library which reflects a C₄ transcriptome read library will be generated. This method does not take any 5’ or 3’ bias into account since it is not known whether the Cleome read libraries are indeed biased. The script used in this study is given in Supplementary Fig. S3 available at JXB online. In a second step, sequence variation was introduced. The most common sequencing error with 454 technology is miscalled homopolymer stretches; in other words, if the same nucleotide occurs multiple times in a row, the software may miss or add a nucleotide to the stretch. A coding sequence, the target of a transcriptome project, very rarely has homopolymer stretches. To determine the pattern of sequence variation, which may have resulted from either genetic variation in the sample or incorrect base calls during sequencing, reads from C. gynandra were mapped onto the C. gynandra unigenes generated in Bräutigam et al. (2011), and for each position nucleotide differences were annotated (Supplementary Fig. S3, three examples shown). The nucleotide substitutions appeared random (Supplementary Fig. S3, three examples shown). Therefore, to generate simulated read libraries with sequence variation present, error rates of 1, 3, and 5% were introduced at random positions in the read library with a Perl script. This script is also available in Supplementary Fig. S3.

**Assembly**

The source code for all assemblers except CLC is publicly available and was downloaded from public repositories. A free CLC trial version was downloaded from the company’s website. All assemblers were run on a Linux Workstation with Dual Core CPU and 8 Gb of RAM. For both SOAP and Velvet, the k-mer size for the assembly was set to 19. MIRA was started with recommended parameters for expressed sequence tag (EST) assembly, using default settings otherwise: denovo, est, accurate, 454. CAP3 was run with default parameters and therefore an overlap identity requirement of 75% and minimal overlap length of 30 bases.
TGICL is a pipeline consisting of a megablast-like tool, which was run with default parameters (overlap of at least 40 bases and 94% identity) which pre-clusters the reads into bins each of which contains only reads that overlap at least partially. After clustering, CAP3 addresses each cluster separately for assembly (parameters identical to previous assembly). CLC was run with default settings. After the assemblies, assembly parameters were calculated with Perl scripts and Linux commands.

**SNP identification**

Reads were aligned to the reference contig sequences produced by CLC using proprietary tools integrated into the CLC genomics workbench software suite, and an SNP was called if at least six reads covered the position, if at least two different nucleotides were present with at least two different reads each, and if 40% of the detected variation was one of the nucleotides. After automatic detection, the list was manually curated to include only SNPs with frequencies between 30% and 70%.

**Mapping to Arabidopsis and quantification**

Contigs from databases were mapped to *Arabidopsis* in protein space with BlastX. Annotation parameters were extracted using Perl scripts and Linux commands.

**Results and Discussion**

**Determination of the most suitable assembly program**

To produce the best assembly possible with non-normalized read libraries from the C₄ plant *C. gynandra* and its C₃ relative *C. spinosa*, different assembly programs were tested. If the assemblers are used with reads that have no reference genome available as a gold standard; that is, the *Cleome* reads, it is impossible to test the assemblies efficiently. Hybrid contigs, which are assembled from two different transcripts, and transcript coverage cannot be assessed since the transcripts from which the reads originate are unknown. To overcome this difficulty, four artificial read libraries were created. To that end, the quantitative results from Bräutigam et al. (2011) were used to produce a simulated read database from 11,889 *Arabidopsis* transcripts as described in detail in the Materials and methods. In a second step, 'errors', namely random base changes, were introduced at levels of 1, 3, and 5% to mimic both sequencing errors and genetic variation in the libraries. The artificial read libraries were assembled with the open source programs SOAPdenovo (SOAP), Velvet, MIRA, CAP3, and TGICL (Table 1) as well as the commercial program CLC bio genomics workbench (CLC bio, Aarhus, Denmark). Three different measures were used to test the assembly programs. The reduction in total sequence length of the initial libraries compared with the assembled contigs is one measure to test for reduced redundancy. A second measure is the number of contigs which are returned by the assembly. The length distribution is tested with the N25 and N50. If the contigs are sorted by length and one checks contigs until 25% of the sequence information (i.e. the total base count) is covered, the N25 is the length of the shortest contig in the list. In analogy, the N50 is the length of the shortest contig if one carries on until 50% of all sequence information is contained. With the artificial library and without sequencing errors, all assemblers reduce the redundancy by ~90%, indicating that a large proportion of reads are assembled into contigs. The programs produce between 27,315 contigs for SOAP and 12,277 and 11,787 contigs for TGICL and CLC, respectively, with N50s between 476 nucleotides for SOAP and 732 nucleotides for CLC (Table 1). Since 2128 of the transcripts are only represented by one read (Bräutigam et al., 2011), all programs retain reads as singletons. The difference in contig number shows that not all assemblers come close to reducing the contig number back to the number of 11,889 transcripts from which reads were drawn. If SOAP was used, the number of transcripts from which the reads were
generated was overestimated at least 2.3-fold, whereas, if CLC was used, the number would be close to correct. In contrast to artificial reads, ‘real world’ sequencing data are never perfect as sequencing instruments make errors and natural populations contain genetic variation. The introduction of an error rate of 1% into the artificial read library causes a drop in redundancy reduction in SOAP to 60% while the other programs maintain numbers comparable with the assembly of the perfect library (Table 1). Both Velvet and MIRA as well as SOAP have dropped the N50 from between 434 nucleotides and 604 nucleotides to between 39 nucleotides and 413 nucleotides, while CAP3, TGICL, and CLC overall produce assemblies very similar to that of the perfect library. The number of contigs generated remains similar for CLC, TGICL, and CAP3, while it doubles in MIRA, triples in Velvet, and is 20-fold in SOAP (Table 1). As these artificial reads are closer to reality, depending on the assembler, the number of transcripts would be estimated as close to correct if TGICL, CAP3, or CLC were used, to a 60-fold overestimate if SOAP was used. The introduction of an even higher rate of 3% variation exacerbates the problems of SOAP, Velvet, and MIRA with losses in redundancy reduction and shorter contigs. At an error rate of 3% in the library, CAP3 starts to lose some reduction in redundancy and has marked losses in long contigs, with N50 dropping from 616 bases at 1% to 458 bases at 3%. Both TGICL and CLC have minor losses in contig length, with the N50s dropping from 678 nucleotides and 718 nucleotides (at 1%) to 611 nucleotides and 689 nucleotides (at 3%). Contig numbers increase slightly for these programs (Table 1). Finally, at an error rate of 5%, all programs are unable to assemble contigs efficiently from the artificial read library (Table 1). The rate of sequence variation in real world data is currently not known. The rate is the sum of the genetic variation in the sampled population and the error rate of transcriptome sequencing.

It is a naïve assumption that long contigs mean ‘best’ assembly. It is critical that the assemblers do not produce hybrid contigs that have joined two different genes together. For the perfect artificial read library, the number of hybrids produced by each assembler was tested: the number of hybrids produced by each assembler was tested: the number of hybrids is ~190 for MIRA, CAP3, TGICL, and CLC, while SOAP produces 434 hybrids and Velvet only produces 29 (Table 1). While TGICL and CLC produce more hybrids than Velvet, they vastly exceed Velvet’s abilities in assembling long contigs. Based on the criteria of contig length distribution and reduction in redundancy (Table 1), among those tested in this study, TGICL, CAP3, and CLC are the most suitable assemblers for non-model transcriptome data. TGICL and CLC are particularly resistant to sequence variation (Table 1).

To learn more about the reason why different assemblers produce such different results (Table 1), for each transcript, the number of reads drawn from the transcripts was plotted against the coverage achieved by the assemblers. The points representing those transcripts with a high number of reads drawn should form a line at the 100% coverage mark, while transcripts with fewer reads drawn may be distributed along the coverage gradient based on transcript length. For the CAP3-based assembly, the TCICL-based assembly, and the CLC-based assembly, the points form a line at the 100% coverage mark while points representing transcripts from which fewer reads were drawn are distributed along the coverage gradient (Fig. 1). However, for the assemblies performed with MIRA, Velvet, and SOAP, the points which represent transcripts from which many reads were drawn form a cloud above those from which few transcripts were
Table 2. Assembly results of the C. gynandra library with five different assemblers and the results of the CLC assembly of the C. spinosa read library

For C. gynandra 368,333 reads with 85,681,233 bases and for C. spinosa 284,318 reads with 65,525,139 bases were assembled. Remaining sequence length is the number of bases in the contigs after the assembly compared with the number of bases in the original sequence reads.

<table>
<thead>
<tr>
<th></th>
<th>C. gynandra</th>
<th>Velvet</th>
<th>MIRA</th>
<th>CAP3</th>
<th>TGI CL</th>
<th>CLC</th>
<th>C. spinosa</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of contigs</td>
<td>383,907</td>
<td>92,149</td>
<td>30,785</td>
<td>20,259</td>
<td>19,019</td>
<td>17,851</td>
<td>16,770</td>
</tr>
<tr>
<td>N25</td>
<td>245</td>
<td>288</td>
<td>719</td>
<td>821</td>
<td>885</td>
<td>968</td>
<td>859</td>
</tr>
<tr>
<td>N50</td>
<td>106</td>
<td>173</td>
<td>434</td>
<td>496</td>
<td>529</td>
<td>596</td>
<td>521</td>
</tr>
<tr>
<td>N75</td>
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<td>84</td>
<td>290</td>
<td>344</td>
<td>357</td>
<td>379</td>
<td>337</td>
</tr>
</tbody>
</table>

Fig. 2. Venn diagram of the mapping results of the contigs from C. spinosa and C. gynandra libraries. After the assembly of the C. gynandra and C. spinosa reads, for each contig the corresponding Arabidopsis transcript was determined. A total of 6211 Arabidopsis transcripts were matched by contigs from both species.

has not been released. Taken together, all the results of the assembler tests point to TGI CL as the oldest but most efficient open source program while the commercial program CLC produces results comparable with or better than TGI CL at a much quicker pace.

Assembling the Cleome transcriptomes

After determining the best assembly program in terms of producing long contigs, few hybrids, and capability of assembling high coverage contigs, the results were confirmed with the C. gynandra read database. Only contig length and reduction in redundancy can be assessed, but not the number of hybrid contigs and the transcript coverage, since the true transcriptome of C. gynandra is unknown. The results of the C. gynandra library assembly mirror those of the artificial library assembly. SOAP produces the largest number of contigs with the smallest N50, while both TGI CL and CLC produce the best results (Table 2). The contig number varies between ~18,000 and ~400,000, and the reduction in redundancy is between 74% and 89% (Table 2). CLC produces the longest contigs, with an N50 of 596 bases compared with TGI CL’s second best of 529 bases. Since CLC is even more capable than TGI CL with the original C. gynandra library, it was also used to assemble the sequence read library from C. spinosa. The number of hybrid contigs cannot be determined. However,
CLC produced 1.6% hybrid contigs with the artificial library (Table 1). In consequence, 1.6% hybrid contigs were considered as the lowest estimate for hybrids in the C. gynandra assembly. The new assemblies generated with CLC (Table 2) are better than the initial assemblies (benchmarks from Brautigam et al., 2011). For example, the N50 increased from 509 bases to 596 bases. When mapping the C. gynandra reads back to the contigs, only 6.5% or 24 068 of the 368 333 reads did not match a contig and thus represent singletons. From C. spinosa, only 7.7% or 21 870 of the 284 318 reads did not match a contig. A second strategy apart from using different assemblers to optimize transcriptome assemblies has been published recently. This work is based on combining two assemblers and producing an assembly of assemblies in a second step (Kumar and Blaxter, 2010). This strategy was not followed for the Cleome assemblies since the artificial read assemblies pointed to the production of hybrid contigs already in the first pass assembly and this problem was expected to be exacerbated in a second pass assembly.

Both Cleome plants were only partially inbred species. All reads were aligned to the consensus CLC contig sequences and SNPs were called as described in the Materials and methods. Based on these stringent criteria, 2323 SNPs were detected in the contigs of C. spinosa, of which eight are complex SNPs with three different possible nucleotides, and 2367 SNPs were detected in the contigs of C. gynandra, of which seven are complex SNPs. The SNPs annotated with this publication do not contain deletions or insertions of nucleotides and are thus unaffected by 454s technology’s inability to read homopolymer stretches correctly. The SNP detection tables are available with this publication (Supplementary Table S1 at JXB online). Short read technologies such as Illumina, SOLiD, or HeliScope could be used to extend the SNP list by sampling and sequencing more individuals. Based on the SNP profile in the transcribed sequences alone, both Cleome species could be developed into inbred lines, of which one could be used for mutagenesis and the other for providing markers to map the mutation.

Comparison of the Cleome assemblies

The final assemblies from both Cleome species were compared with each other. Of the contigs, 86% and 87% could be annotated with A. thaliana using BlastX with a cut-off value of e^{-4}. A similar percentage of reads could be mapped to the Arabidopsis transcriptome (Brautigam et al., 2011). While C. gynandra contigs mapped to 9203 unique Arabidopsis transcripts, C. spinosa contigs mapped to 8598 unique Arabidopsis transcripts. Assuming a plant has ~30 000–40 000 protein-coding genes (Swarbreck et al., 2008; Paterson et al., 2009; Schnable et al., 2009) and about half of these genes are expressed in leaves (Schmid et al., 2005), one would expect ~15 000–20 000 transcripts expressed in leaf tissue of a plant. While the number of contigs is well within the estimated number of transcripts, the number of Arabidopsis transcripts matching the Cleome transcripts clearly is not. Two conclusions can be drawn based on this observation. First, the coverage of leaf transcripts is probably not complete. Even if the species Cleome had more genes (i.e. due to a genome duplication event), the genes expressed in leaves should roughly match half of the Arabidopsis transcripts, or ~15 000. Secondly, the number of contigs exceeds the number of matching Arabidopsis transcripts by a factor of ~2. The contig assembly thus probably retains an ~2-fold redundancy. This redundancy is probably due both to imperfect assemblies, for example due to sequence variation introduced by >2000 SNPs in the species (Supplementary Table S1 at JXB online), and to non-overlapping sequence reads, given that

![Fig. 3](attachment:image-url)
the reads libraries are highly dynamic, spanning five orders of magnitude (Bräutigam et al., 2011).

Of 9203 and 8598 contigs, in C. spinosa and C. gynandra, respectively, roughly two-thirds or 6211 were shared among both species, and 2980 and 2373 were unique to either species (Fig. 2). When only reads and not contigs are mapped to Arabidopsis, about half of the transcripts in the reference are identified (Bräutigam et al., 2011) compared with about a third with contig mapping. There are two possible explanations. On the one hand, reads are shorter than contigs, so the mapping may not have been as precise and more different Arabidopsis transcripts were tagged. On the other hand, the reads were mapped to a minimal genome devoid of transcripts resulting from a whole-genome duplication and subsequent tandem duplications (Bräutigam et al., 2011). Assuming equal mapping accuracies of reads and contigs, one may argue that the Cleome species lack a larger proportion of transcripts resulting from duplications.

To study the end-point of differentiation into C_4 leaf tissue, mature leaves were sampled from a C_3 and a C_4 species and the sequence libraries were not normalized prior to sequencing. These strategic decisions limit the qualitative sequence information: the contig database represents only transcripts expressed in mature leaf tissue and the depth of sequencing is relatively shallow, compared with normalized libraries. Read mapping to Arabidopsis indicated that primary metabolism as well as categories related to leaf functions were well represented in the Cleome libraries, while categories such as regulation were under-represented (Bräutigam et al., 2011). Representation was tested using the contigs as well (Fig. 3). The results essentially mirror those obtained with reads alone (Fig. 3; Bräutigam et al., 2011) The use of only mature leaf tissue clearly limits the number of transcripts which can be identified and it hinders the even distribution throughout functional categories. The sequencing of additional libraries from developing leaves and/or other plant tissues will increase both the number of transcripts and their coverage, as well as evening out the category representation. Based on the limitations in the contig database the question arises as to what degree the transcriptomes can be used to study the end-point of differentiation to a fully mature C_4 leaf.

To answer this question, the transcript coverage for transcripts known to be involved in C_4 photosynthesis was extracted and visualized (Fig. 4). Candidate transcripts for...
regulatory processes were randomly chosen from Bräutigam et al. (2011) and also visualized (Fig. 4). By matching transcript sequences from genes overexpressed in C4 tissue (Bräutigam et al., 2011) with their contig sequences (this work), studies at the molecular level can indeed be initiated. For example, the full-length transcripts for the C4 cycle enzymes of C. gynandra as well as the transport protein PPT known to be involved in C4 photosynthesis can be extracted from the C. gynandra contig file (Fig. 4; Supplementary Fig. S1 at JXB online). Additional transcripts probably involved in the C4 cycle such as those of candidate transport proteins like the plastidic BASS1, a member of the bile acid:sodium symporter family, or DIC, a dicarboxylate carrier at the mitochondrial membrane, can be extracted and studied. When the contig length of these transcripts is compared with the length of the Arabidopsis representative transcript model, the contigs achieve >85% coverage. Known regulators of C4 photosynthesis, which are more highly expressed in the C4 species, are PEP kinase (72% coverage), PPDK regulatory protein (108%), and a golden2-like transcription factor (100%). Candidate regulators, such as glucanases, potentially involved in increasing the open probability of plasmodesmata (Bräutigam et al., 2011) are covered to 90% and 94%, respectively. Similar numbers are achieved for candidate transcription factors up-regulated in C4 (Fig. 4). Since known and candidate transcripts for C4-related processes are covered in full or nearly so, the transcriptome databases are suitable to study the end-point of C4 differentiation. There may also be regulatory transcripts which need to be down-regulated in mature C4 tissue and hence less abundant in the transcriptome database. For the two transcription factors down-regulated in the C4 species which were tested, approximately half-length transcripts were assembled, indicating that the coverage is good enough to initiate further analysis. The NGS project for Cleome succeeded in both the comparative quantification of gene expression between a C4 and a C3 species (Bräutigam et al., 2011) and in providing a sequence resource for further research.

Conclusion

The artificial read libraries constructed based on quantitative information enabled the study of different assembly programs and identified two useful assemblers for next-generation mRNA-Seq data: TIGICL and CLC bio genomics workbench. The application of the assemblers to real world data of C. gynandra confirmed the results of artificial library assembly. The contig databases for mature C3 and C4 leaves represent the pathways of mature leaves well and enable the study of the end-point of C4 photosynthetic differentiation.

Supplementary data

Supplementary data are available at JXB online.

Figure S1. The contig database of C. gynandra.
Figure S2. The contig database of C. spinosa.
Figure S3. Scripts for producing the simulated read libraries.
Figure S4. The sequence variation detected in the C. gynandra read library exemplified by three contigs with annotated sequence variation.

Table S1. Single nucleotide polymorphism tables for C. gynandra and C. spinosa.

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